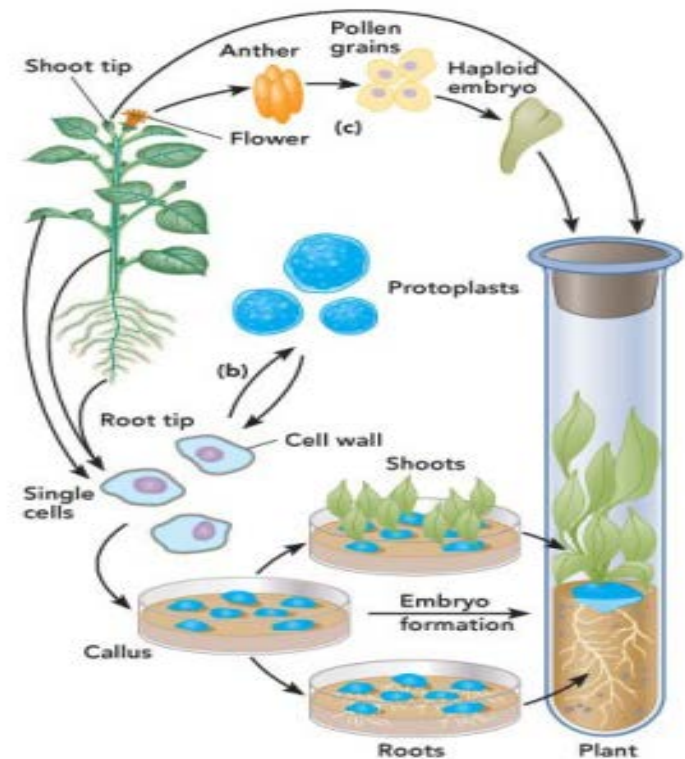
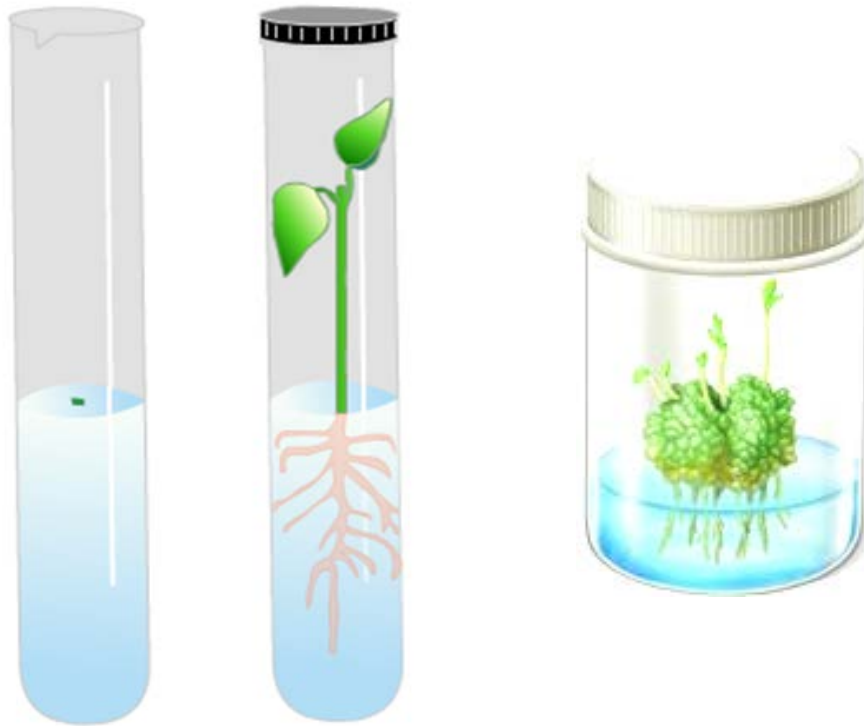


PLANT CELL & TISSUE CULTURE

DR. OMAR SABRY

PLANT CELL & TISSUE CULTURE

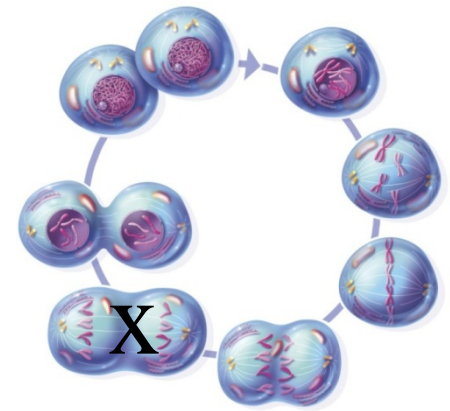
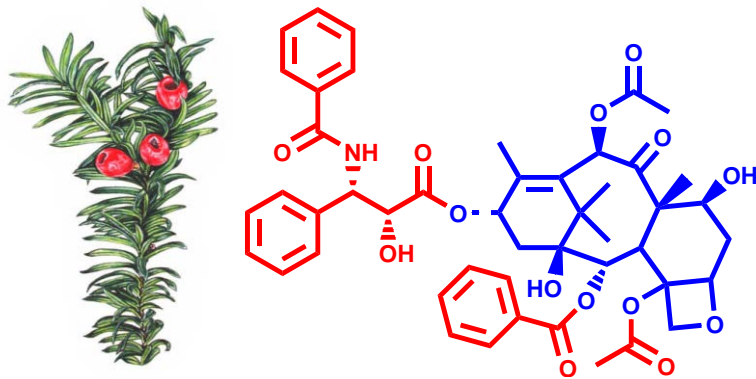
- **In vitro aseptic cultivation of Isolated plant organs or cells (root tips, shoot tips, leaves, embryos , cells on a nutrient medium).**



PLANT CELL & TISSUE CULTURE

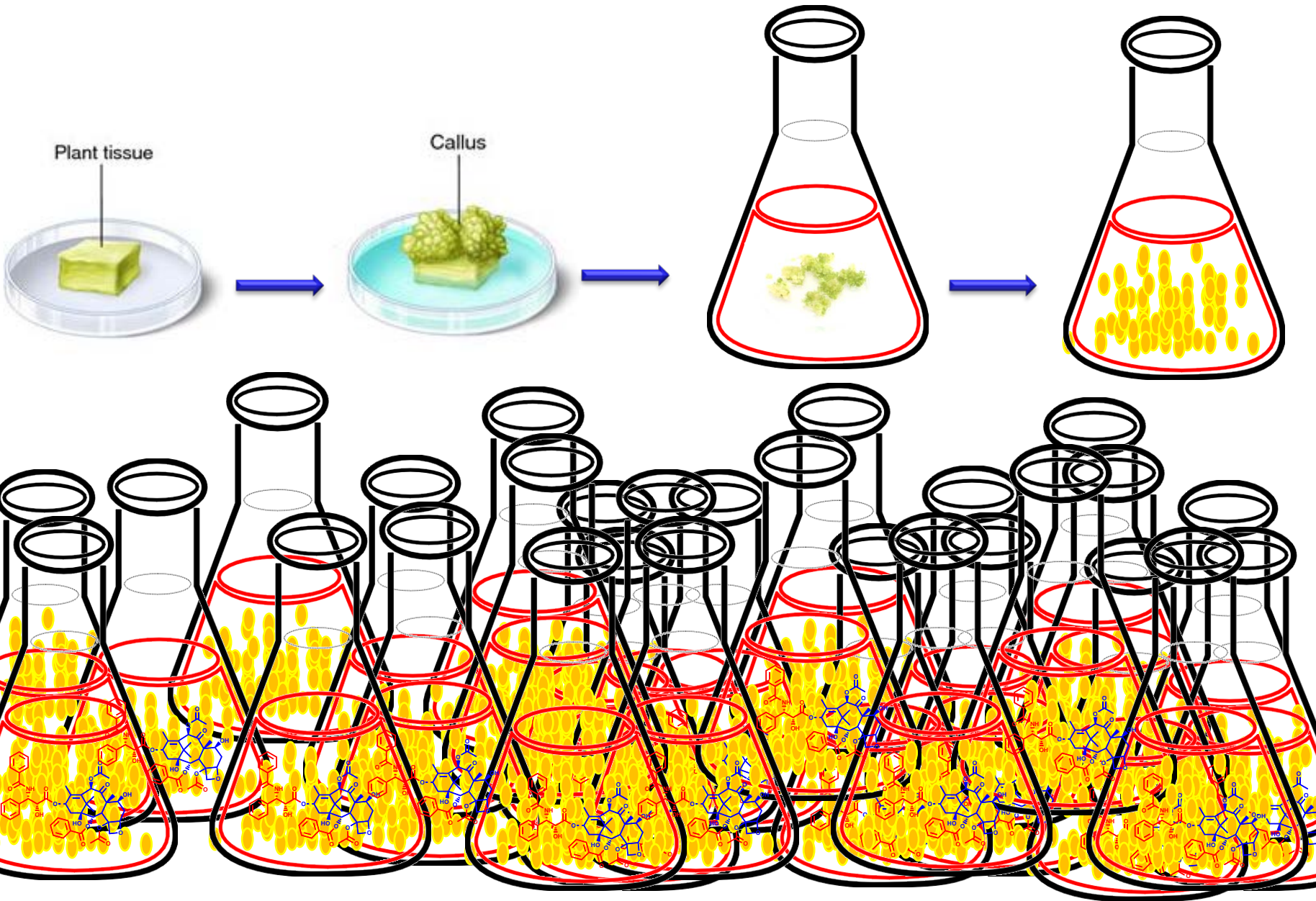
- ❑ Development of commercial production of expensive bio-medicaments (Taxol).**

- ☐ **Anti-cancer from *Taxus brevifolia*.**



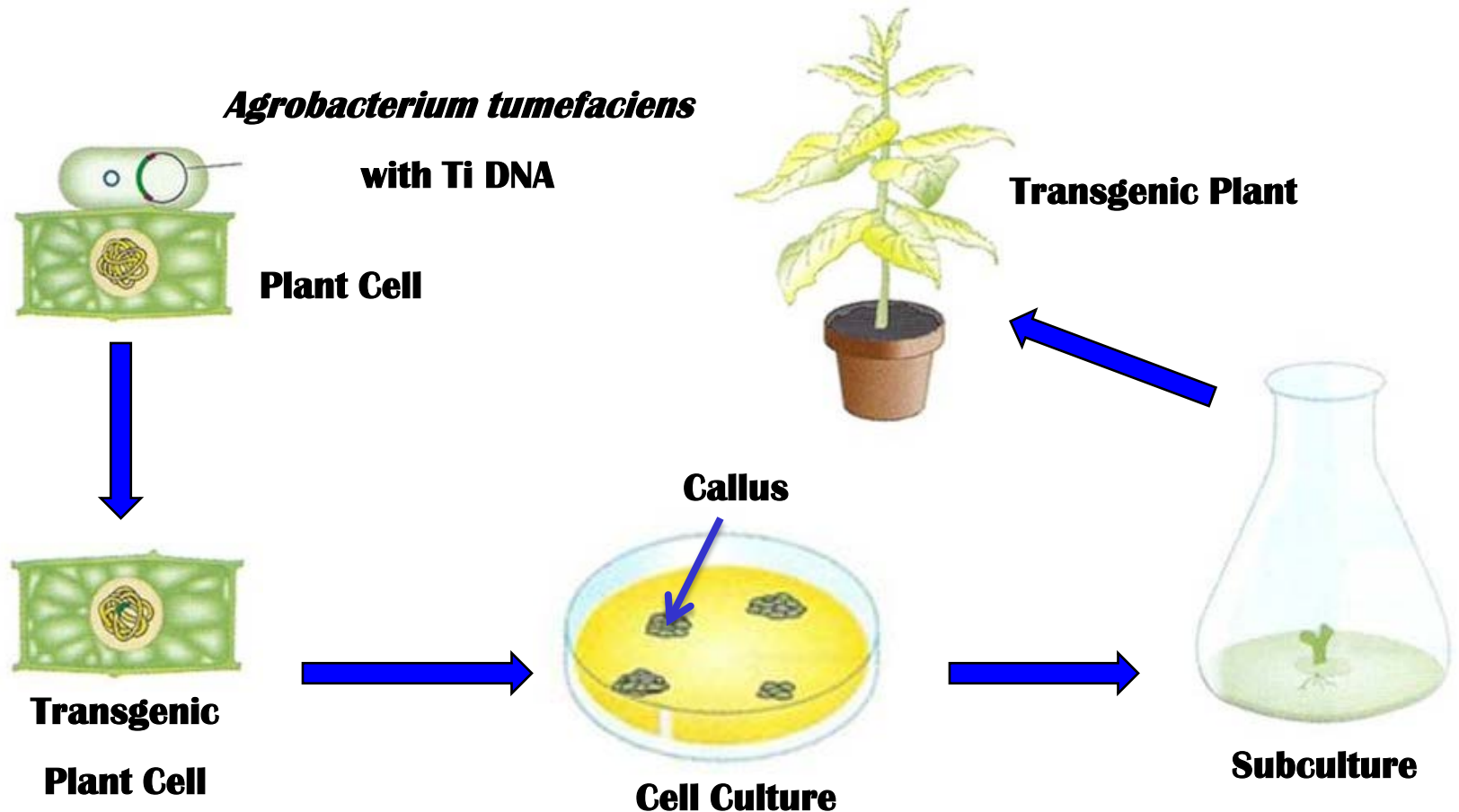
- ☐ **100 years old tree suitable for stem bark**
- ☐ **3 mature trees = 1 gm of Taxol.**
- ☐ **2 gm Taxol for Cancer treatment.**

PLANT CELL & TISSUE CULTURE



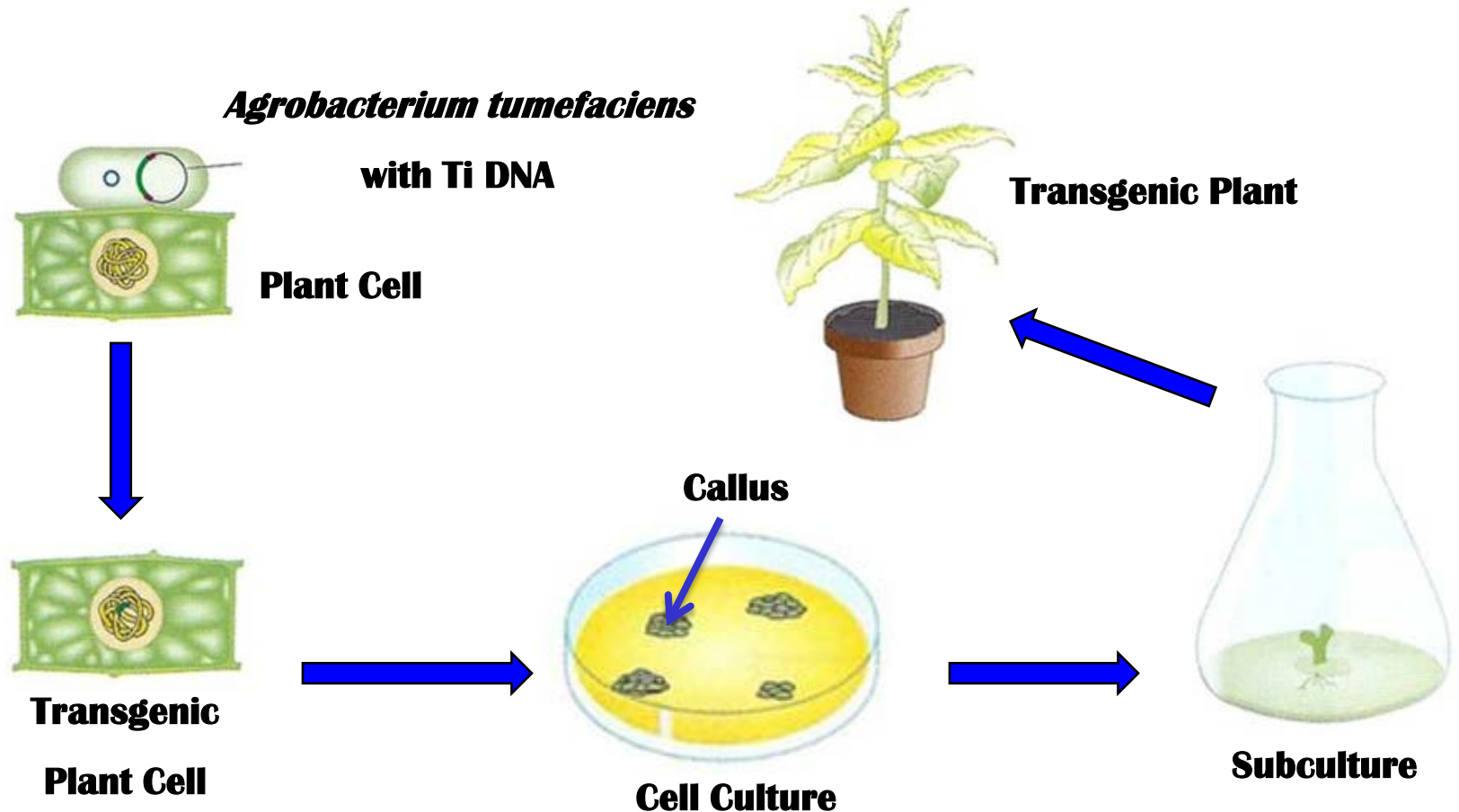
PLANT CELL & TISSUE CULTURE

- ❑ Improvement of medicinal plants by genetic engineering e.g. *Atropa belladonna* gives atropine and morphine.



PLANT CELL & TISSUE CULTURE

- ❑ **Improvement of medicinal plants by genetic engineering e.g. Insects resistant and microbes resistant plants.**



PLANT CELL AND TISSUE CULTURE LABORATORY

(A) Washing area



PLANT CELL AND TISSUE CULTURE LABORATORY

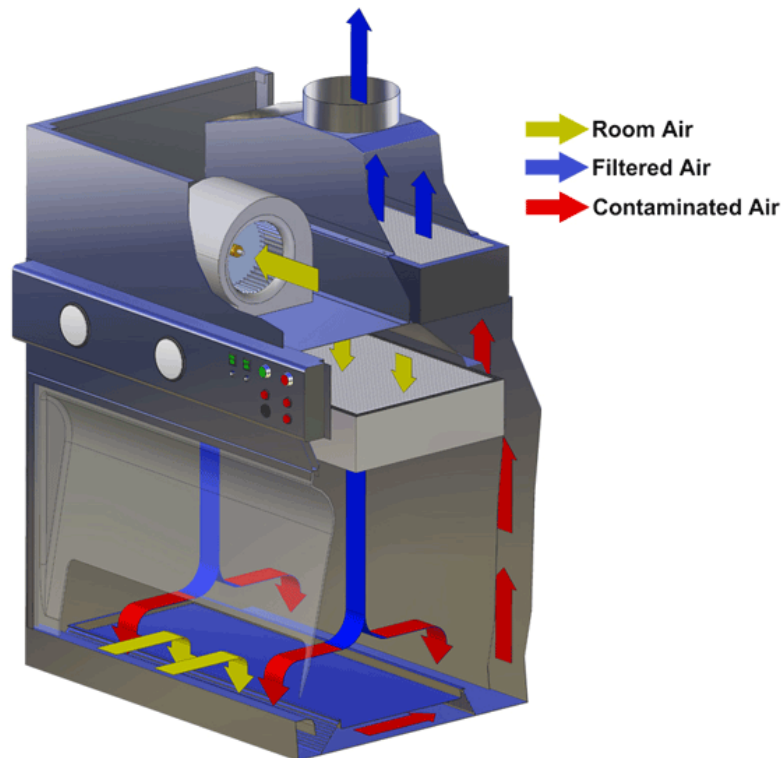
(B) Media preparation room



PLANT CELL AND TISSUE CULTURE LABORATORY

(C) Transfer Area

Laminar air - flow cabinets



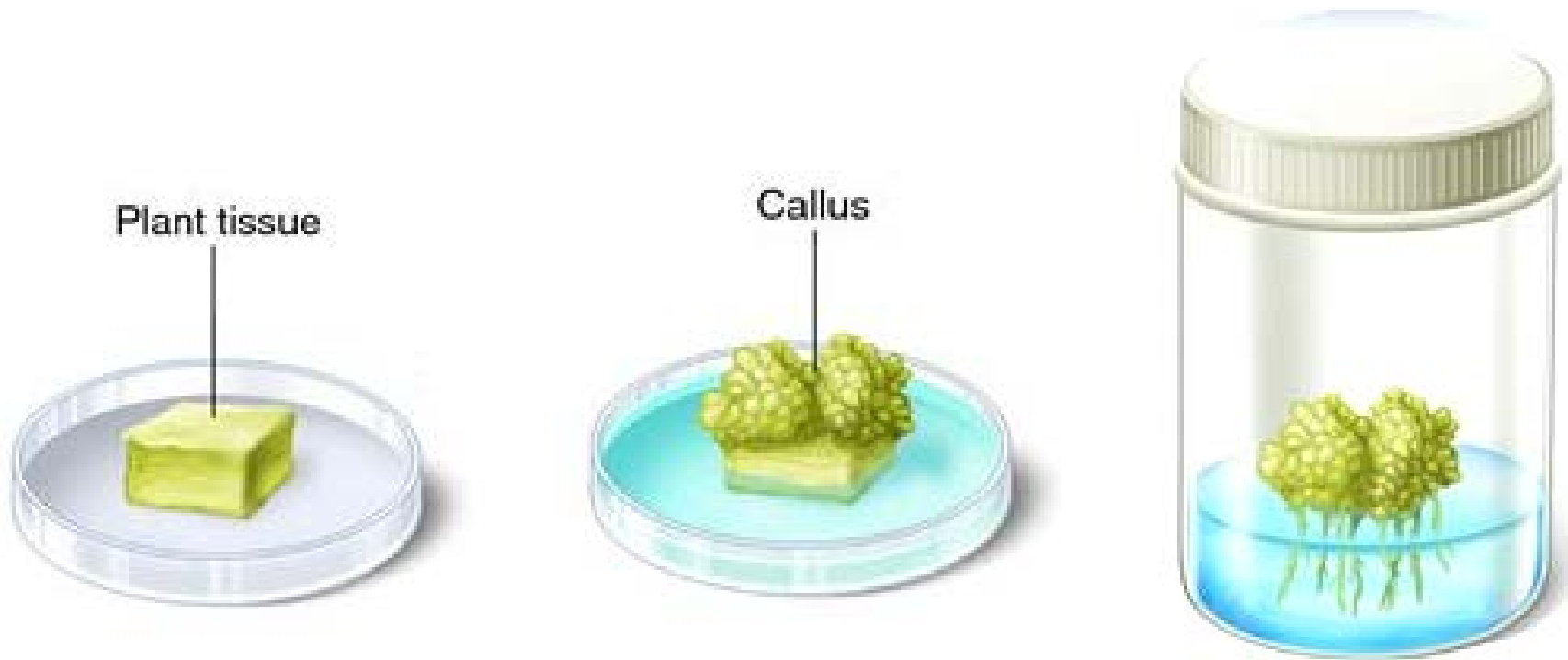
PLANT CELL AND TISSUE CULTURE LABORATORY

(D) Culture room



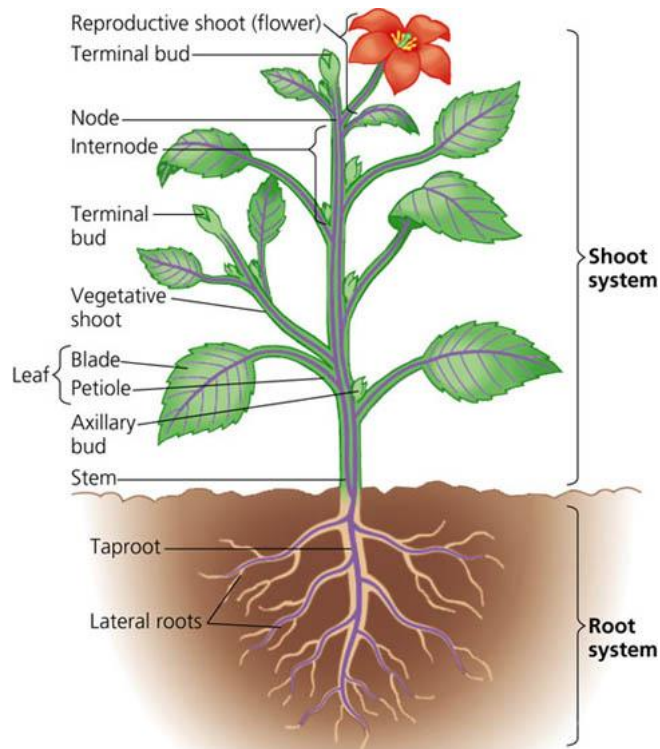
CULTURE VESSELS

- ❑ **Borosilicate or Pyrex glassware should be used.**
- ❑ **Soda glass toxic to tissues with repeated use.**

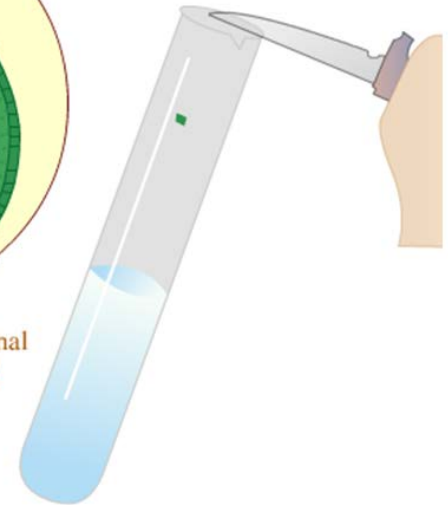


SOURCES OF TISSUE EXPLANTS

- ❑ **Organ with meri-stimatic cells Pith, root tips, shoot tips, embryos, anthers and young leaves.**

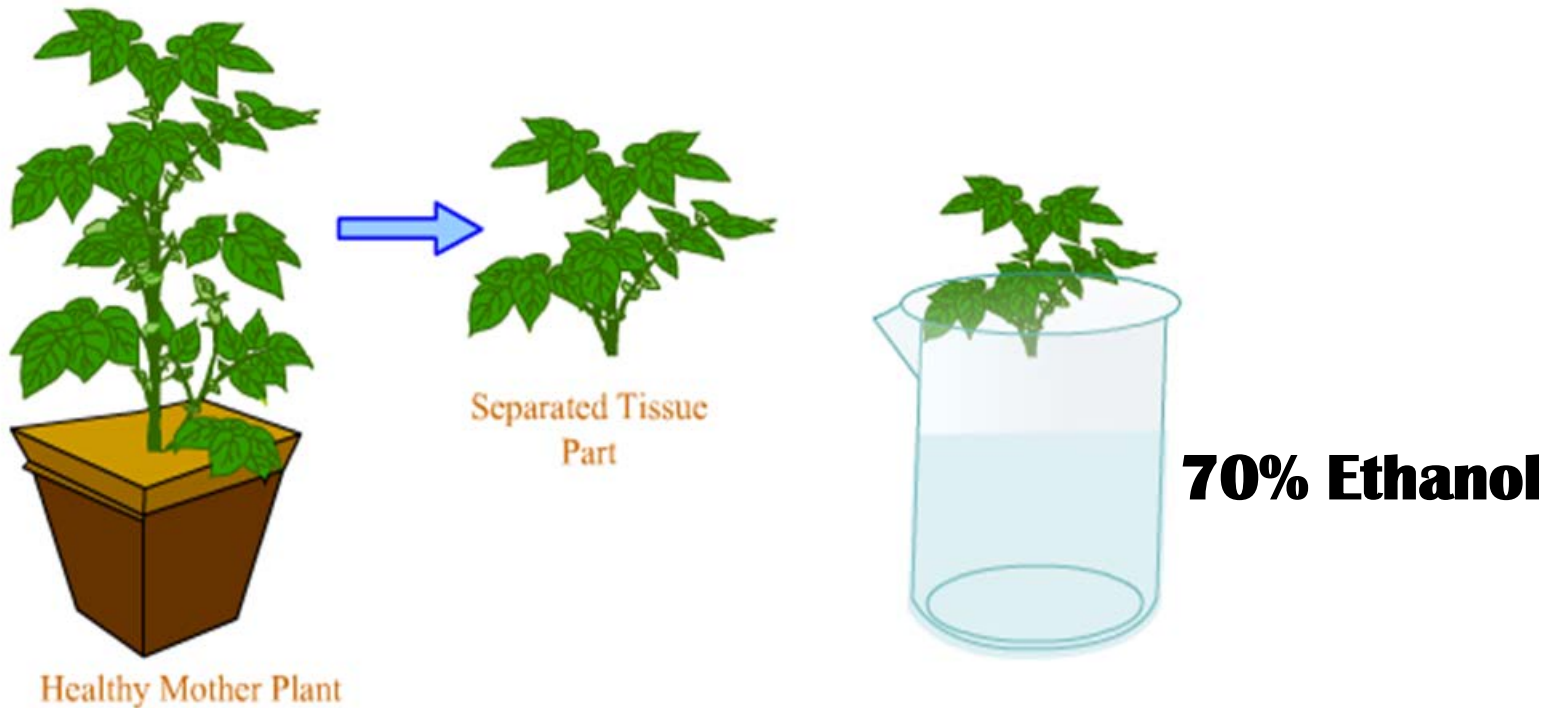


Meristematic tissue of terminal
(microscopic appearance)



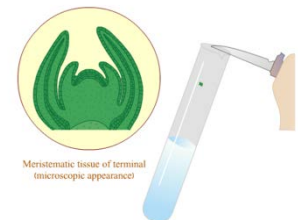
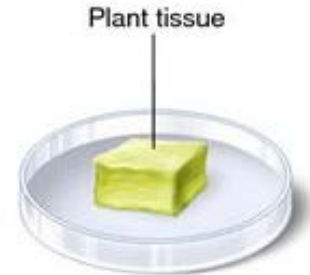
STERILIZATION OF PLANT MATERIAL

- ❑ **Tissues must be surface sterilized before planting on the nutrient medium.**



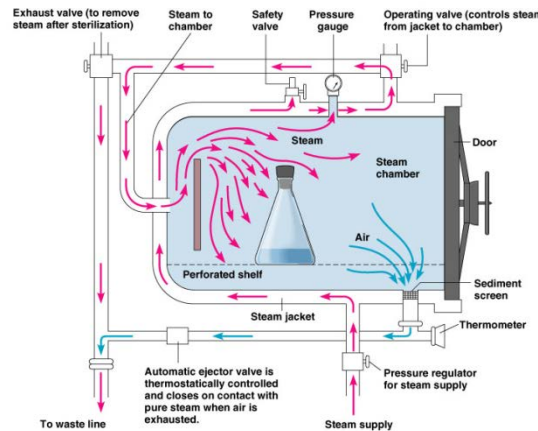
STERILIZING AGENTS USED IN PLANT STERILIZATION

- ☐ **70% ethyl alcohol (No methanol)**
- ☐ **Sodium hypochlorite 25%**
- ☐ **After disinfection, carefully trim by cutting away all bleached and dead tissue .**
- ☐ **Rectangular or cylindrical pieces approximately 5 mm in diameter excised transferred to agar plates containing nutrient medium.**



STERILIZATION OF GLASS WARE, PLASTIC WARE AND OTHER INSTRUMENTS

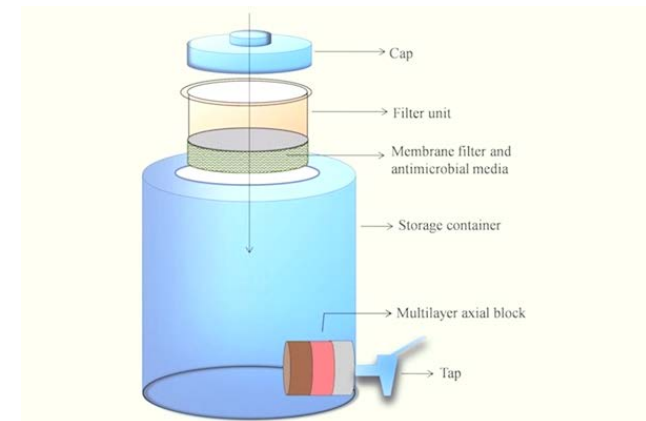
- ❑ Autoclaving or dry heating in an oven at 160 -180°C for 3hr.



- ❑ Plastic ware can be heat sterilized by autoclaving at 121 °C.
- ❑ Forceps, scalpels, needles and spatulae sterilized by 70% or 95 % ethanol followed by flaming and cooling.

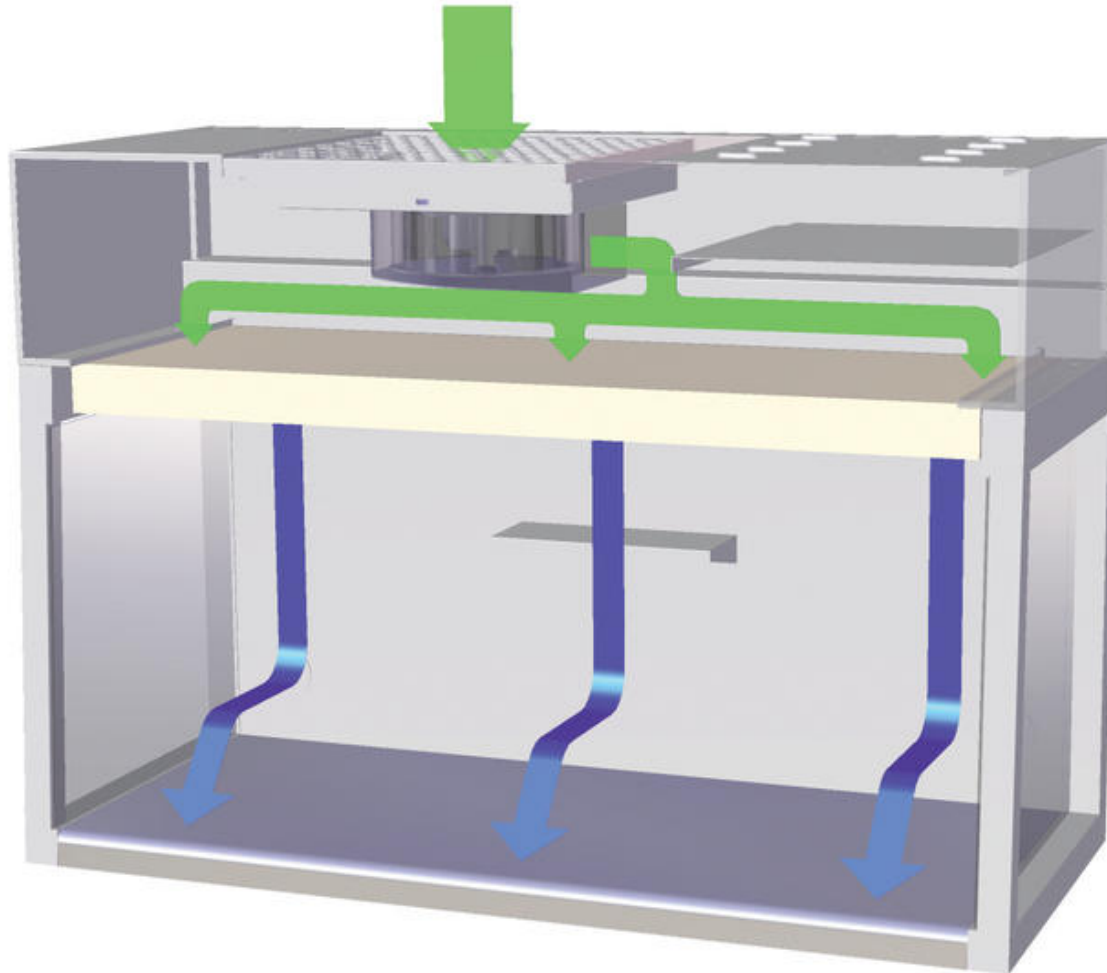
STERILIZATION OF MEDIA

- ❑ **Some growth factors are thermolabile (No autoclaving).**
- ❑ **Medium minus the heatlabile compound autoclaved in a flask kept in the sterilized hood to cool down.**
- ❑ **Solution of the thermolabile compound is sterilized by Micropore membrane filter.**



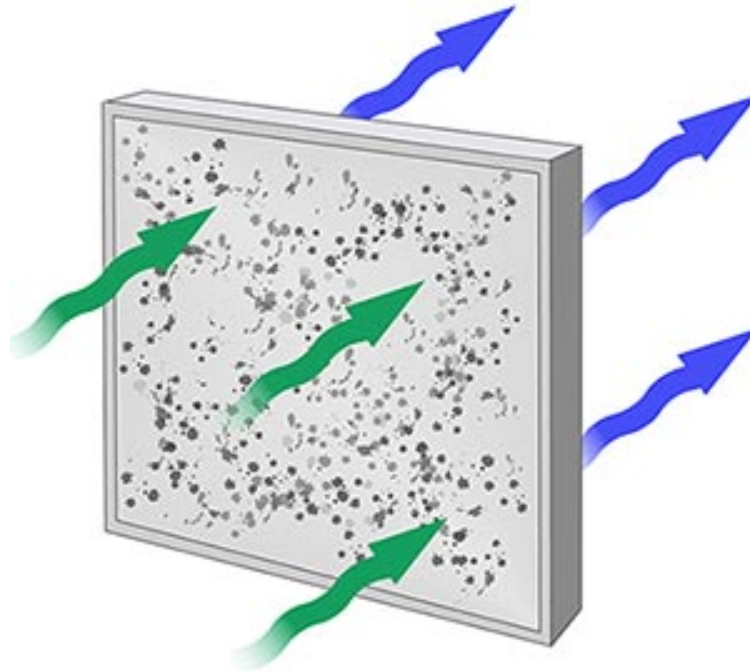
STERILIZATION OF TRANSFER AREA

❑ Laminar airflow cabinets



STERILIZATION OF AIR

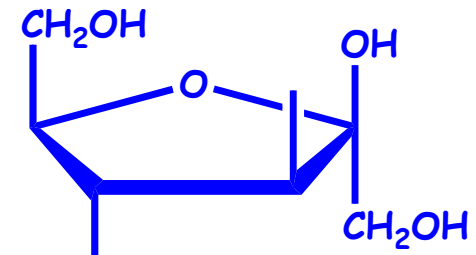
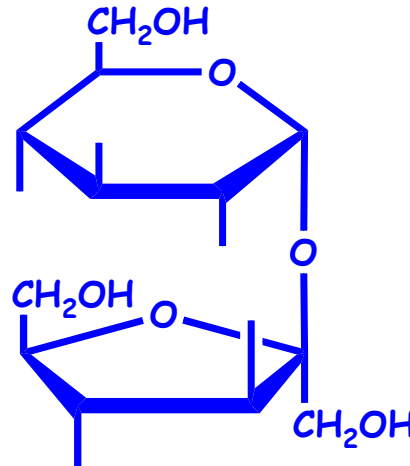
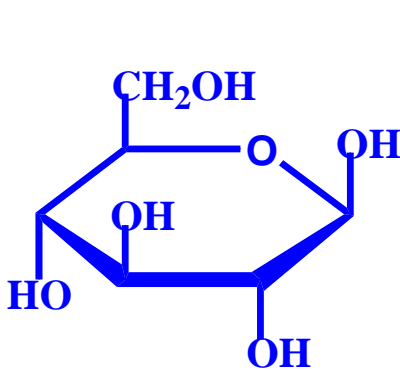
- ❑ **High efficiency particulate air (HEPA) filter**



MEDIA CONSTITUENTS

□ Inorganic Nutrients

- **Macro - or major elements (6): e.g. Nitrogen – Calcium.**
- **Micro - or minor elements (6): e.g. Iron - Cu**



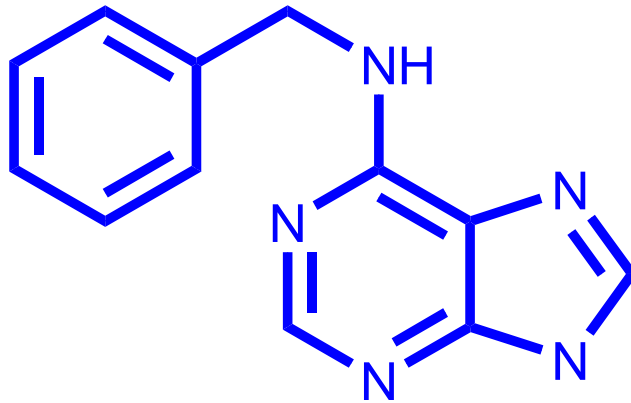
□ Organic Nutrients

- **Nitrogenous substances e.g. Vitamins.
Thiamine - Pyrodoxol**
- **Carbon Source e.g. Sucrose 2-5%, glucose and fructose**

GROWTH HORMONES

CYTOKININS

- ❑ **BAP (benzyl amino purine)**

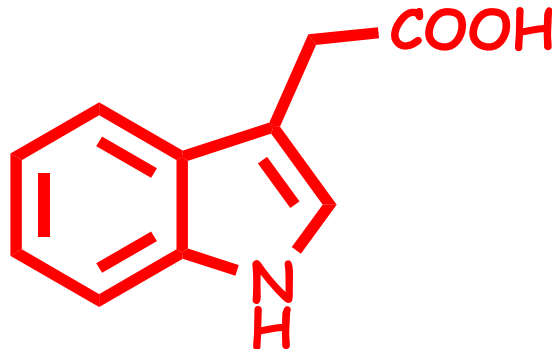


- ❑ **Shoots emerge, Plant rooted with auxin, transferred to potting soil for further growth in the greenhouse as normal plants.**
- ❑ **Equal concentrations gives unorganized fashion**

GROWTH HORMONES

AUXINS

☐ IAA (Indole acetic acid)



☐ **Excess of auxins promotes growth of roots**

☐ **Excess of cytokinins yield shoots.**

AGAR

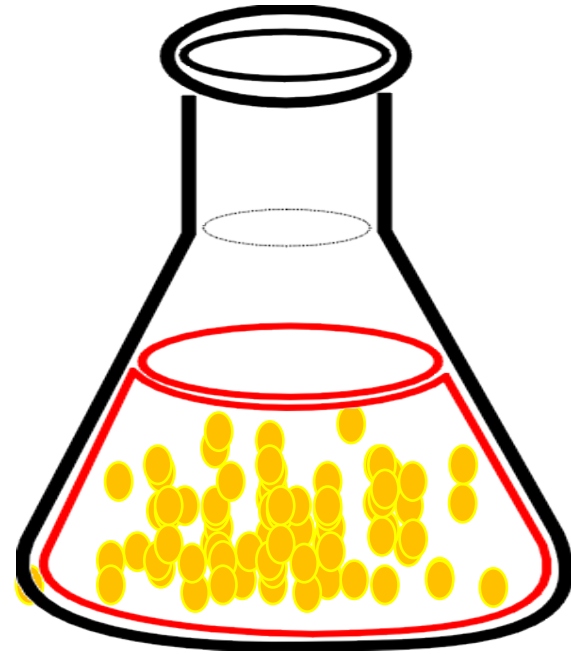
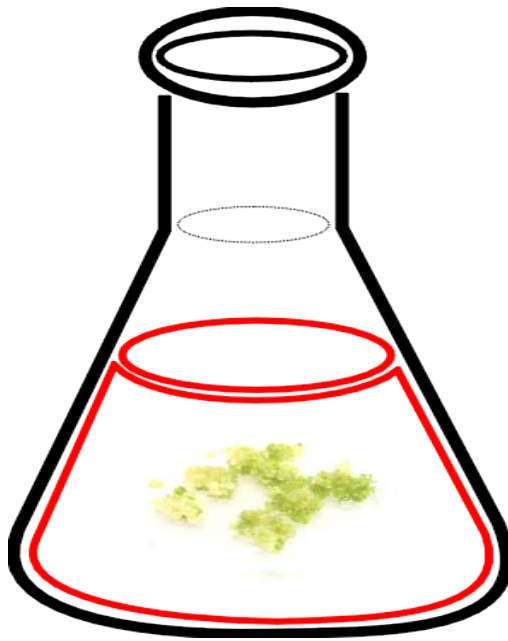
- ❑ **Generally used at a concentration of 0.8 - 1.0%.**
- ❑ **Higher concentration the medium becomes hard not allow the diffusion of nutrients into the tissues.**



- ❑ **Not an essential components of the nutrient medium.**

CELL SUSPENSION CULTURE

- ❑ **Single cells and aggregates can be grown as suspension cultures in liquid medium.**



CELL SUSPENSION CULTURE

- ☐ Regularly aerated either by bubbling sterile air, or gentle agitation.
- ☐ Liquid medium may prove superior to agarified medium.
- ☐ pH of the medium 5.0 - 6.0 before sterilization.



MEDIA SELECTION

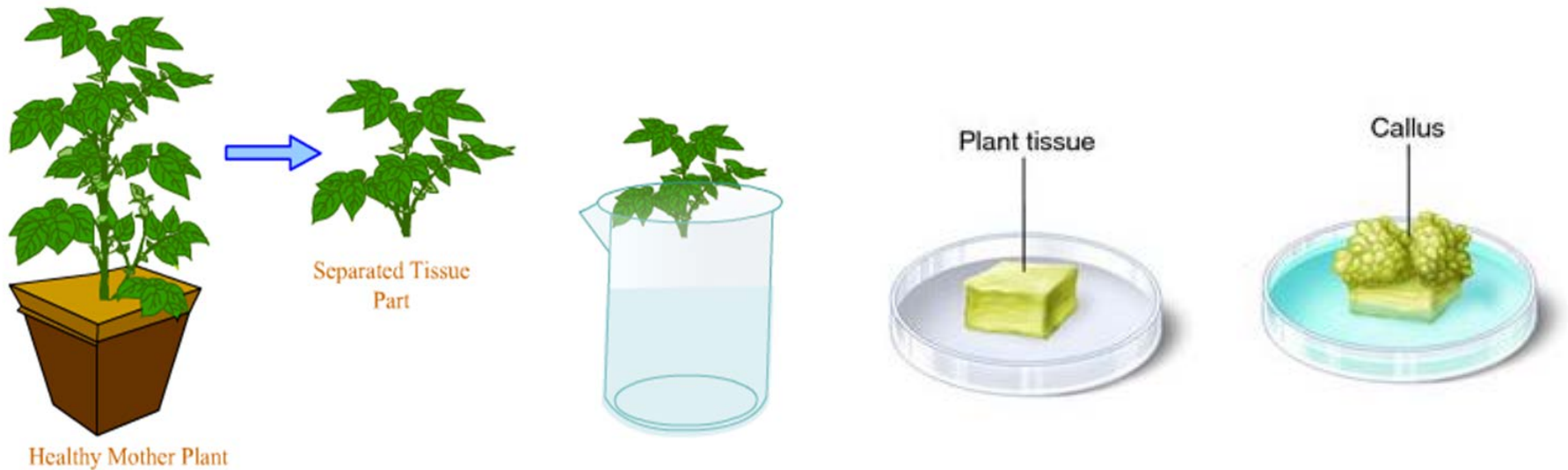
- ☐ **Start with known basal medium.**
- ☐ **Minor qualitative and quantitative changes, through series of experiments a new medium may be evolved.**
- ☐ **Simplest method commercially available dry powdered media, containing inorganic salts, vitamins and amino acids.**



STAGES OF CULTURING

1- PREPARATION OF THE EXPLANT

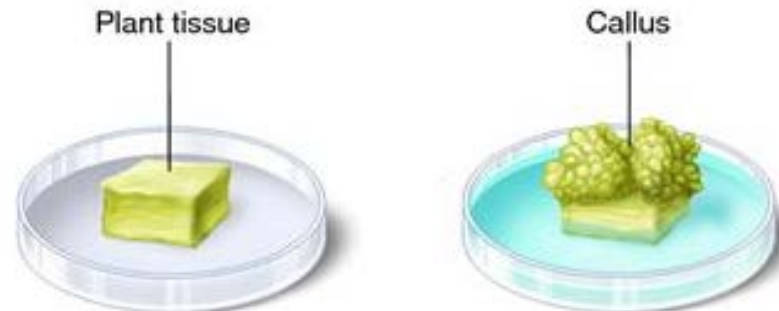
❑ Piece of stem, leaf, or root is disinfected with alcohol or sodium hypochlorite solution and cultured in a specific culture medium.



STAGES OF CULTURING

2- MULTIPLICATION

- ☐ **Number of cells increases.**
- ☐ **Cells cultured from a callus in the presence of low concentration of auxin and cytokinin.**
- ☐ **Calli placed in liquid medium and grown in suspension or placed on a solid substrate to be cultured.**



STAGES OF CULTURING

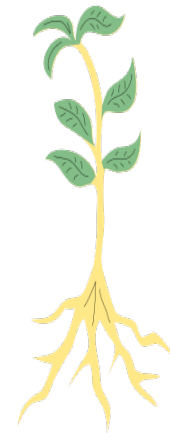
- ❑ **Callus cultures subdivided and re-cultured .**
- ❑ **Exposed to growth hormones, Calli form shoots and roots.**
- ❑ **Callus formation and maintenance must be taken in consideration because when cultured for several weeks any callus show signs of aging.**



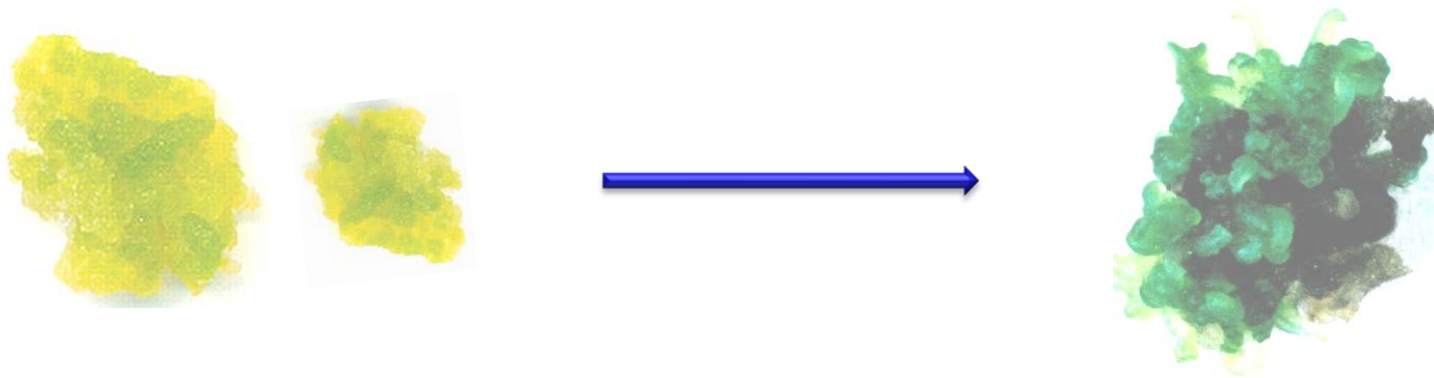
STAGES OF CULTURING

3. PRE-TRANSPLANT STAGE

- ❑ Allows the formation of roots, shoot development, maturation, and development.
- ❑ Cytokinins are reduced or eliminated and auxins are added to promote root development.



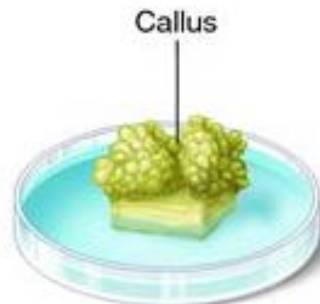
FACTORS INDUCING AGING



- ☐ **Exhaustion of the nutrients.**
- ☐ **Inhibition of nutrient diffusion.**
- ☐ **Accumulation of toxic metabolites.**
- ☐ **Callus tissues periodically transferred to fresh medium (sub-culture) at intervals of 4-6 weeks.**

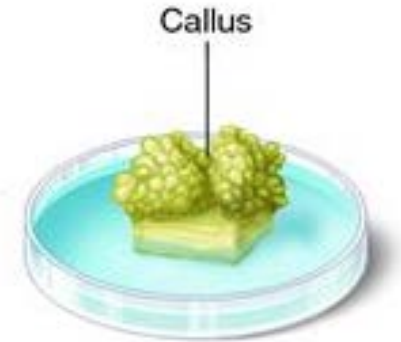
PREPARATION OF CELL SUSPENSION CULTURES

- ❑ **Sections from surface – sterilized plant organs on a nutrient medium containing suitable hormones (auxins and cytokinins).**
- ❑ **The explant exhibits callusing.**
- ❑ **Callus separated from the parent explant transferred to fresh medium.**

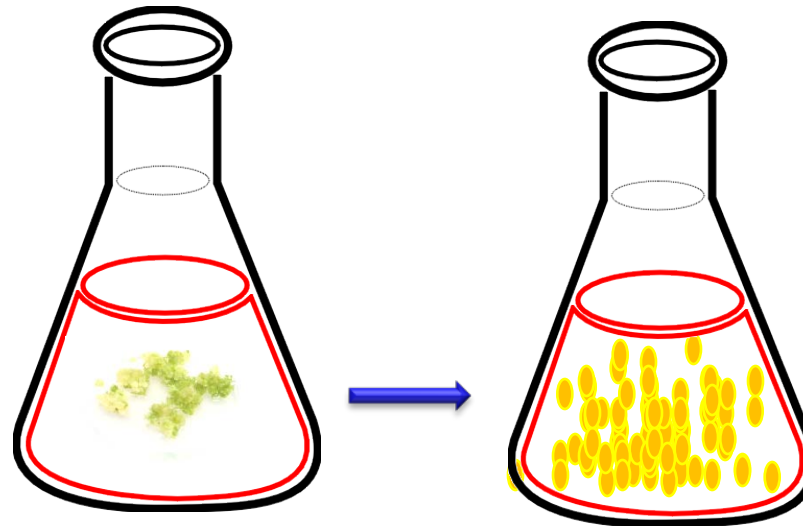


PREPARATION OF CELL SUSPENSION CULTURES

- ❑ Repeated sub-culture on the agar improves friability desirable for raising a fine cell suspension in liquid medium.

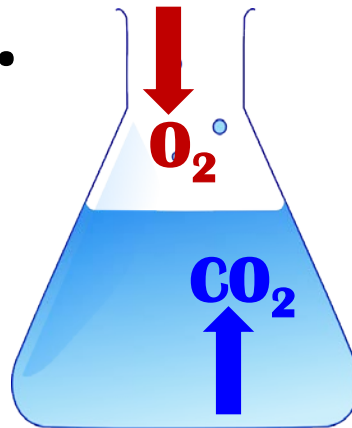


- ❑ On transfer to liquid medium gave fine suspension.



AGITATION FUNCTIONS

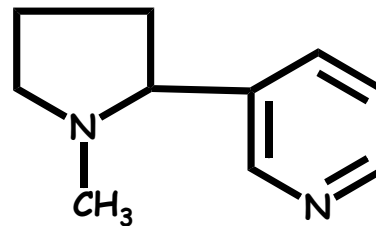
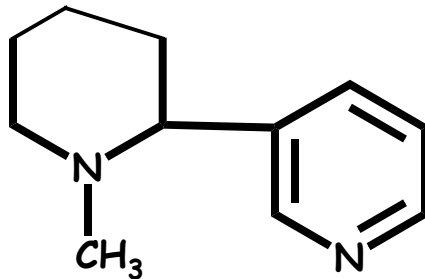
- ❑ **Breaking clumps into single cells.**
- ❑ **Uniform distribution of cells in the medium.**
- ❑ **Gaseous exchange.**



PLANT CELL & TISSUE CULTURE

Why some cell cultures may not produce the natural compounds or produce them in very small amounts?

- ❑ Biosynthesis by cooperation of different organs**
- ❑ Lysine amino acid is converted to anabasine in tobacco roots**
- ❑ followed by the conversion of anabasine to nicotine in leaves.**

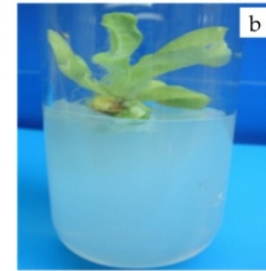
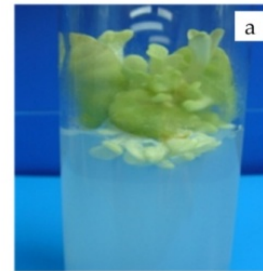


Nicotine



PLANT CELL & TISSUE CULTURE

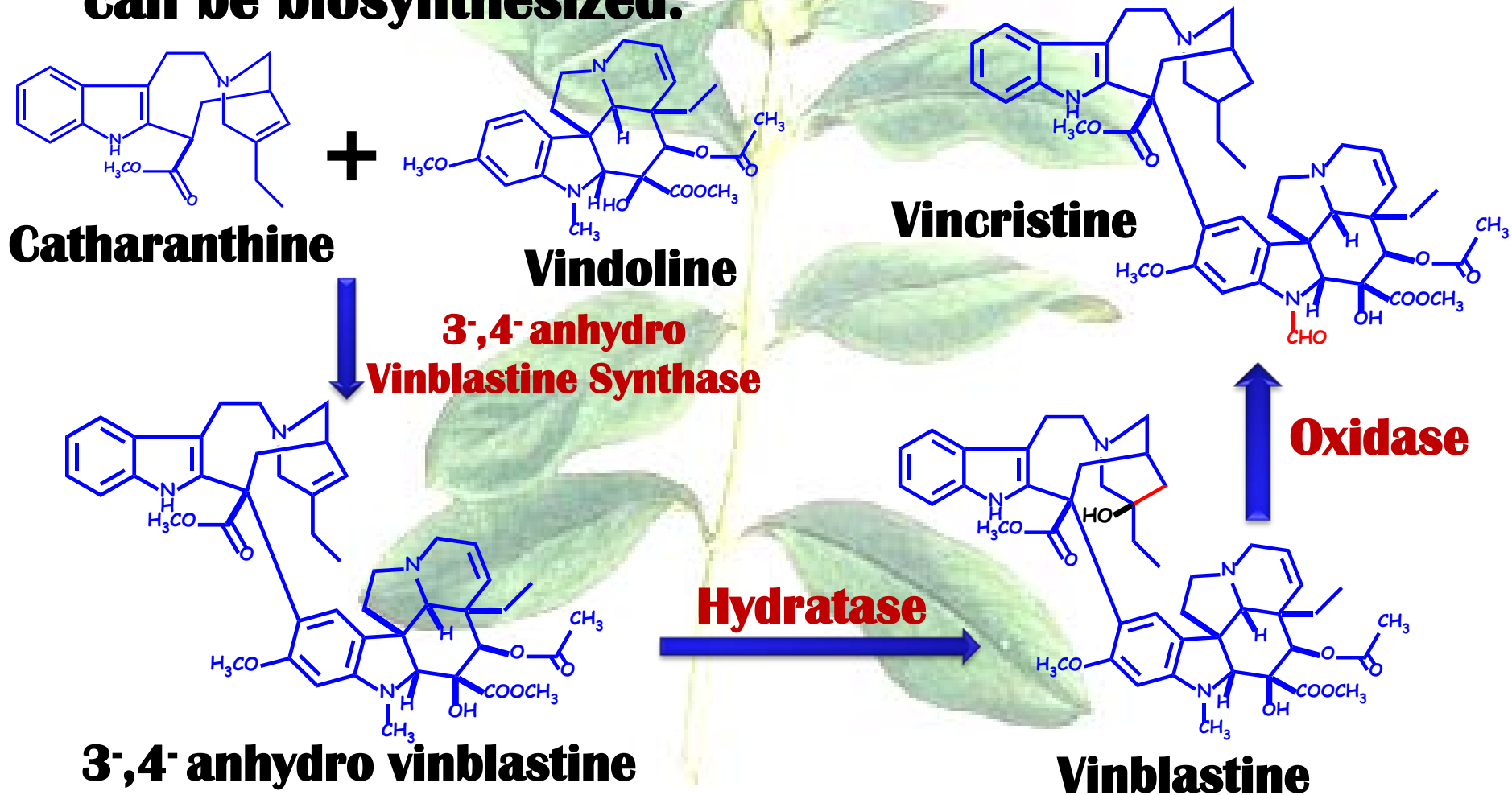
➤ **Callus and shoot cultures of tobacco can produce only trace amounts of nicotine because they lack anabasine.**



❑ **Compound may be produced in only one organ, or in one cell.**

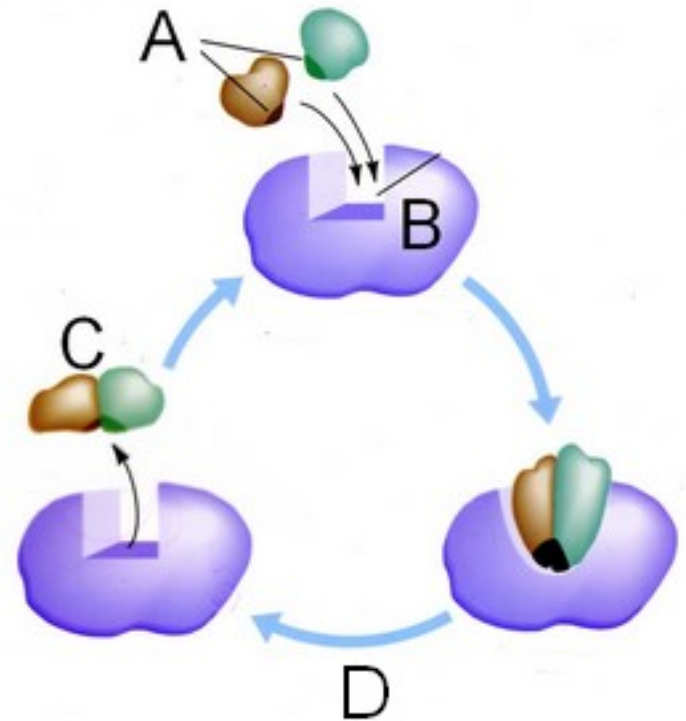
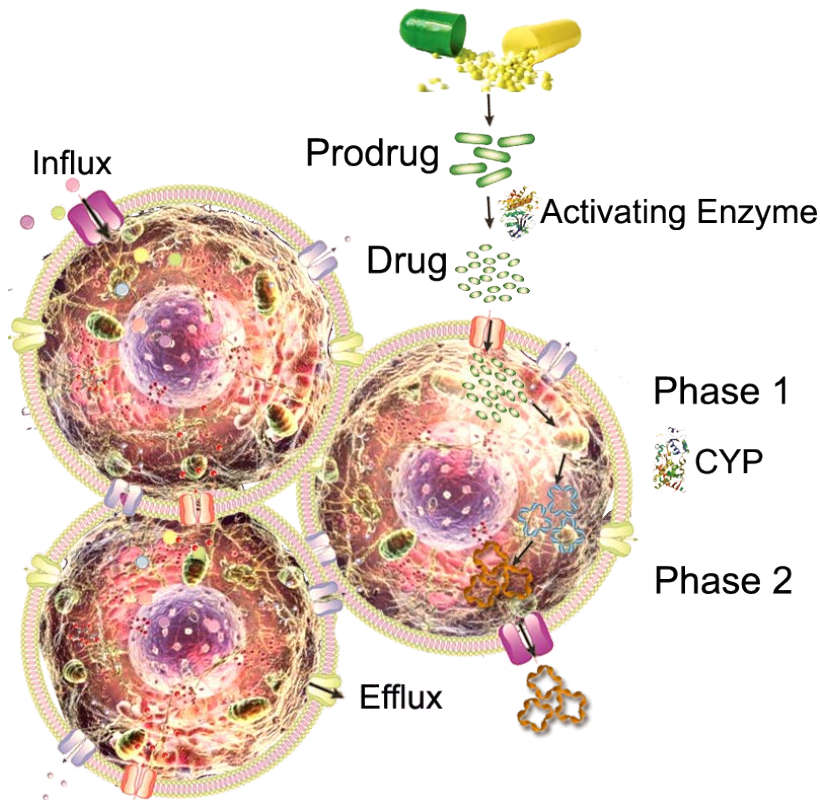
PLANT CELL & TISSUE CULTURE

- ❑ Some degree of differentiation in a cell culture of *Catharanthus roseus* must occur before vincristine can be biosynthesized.



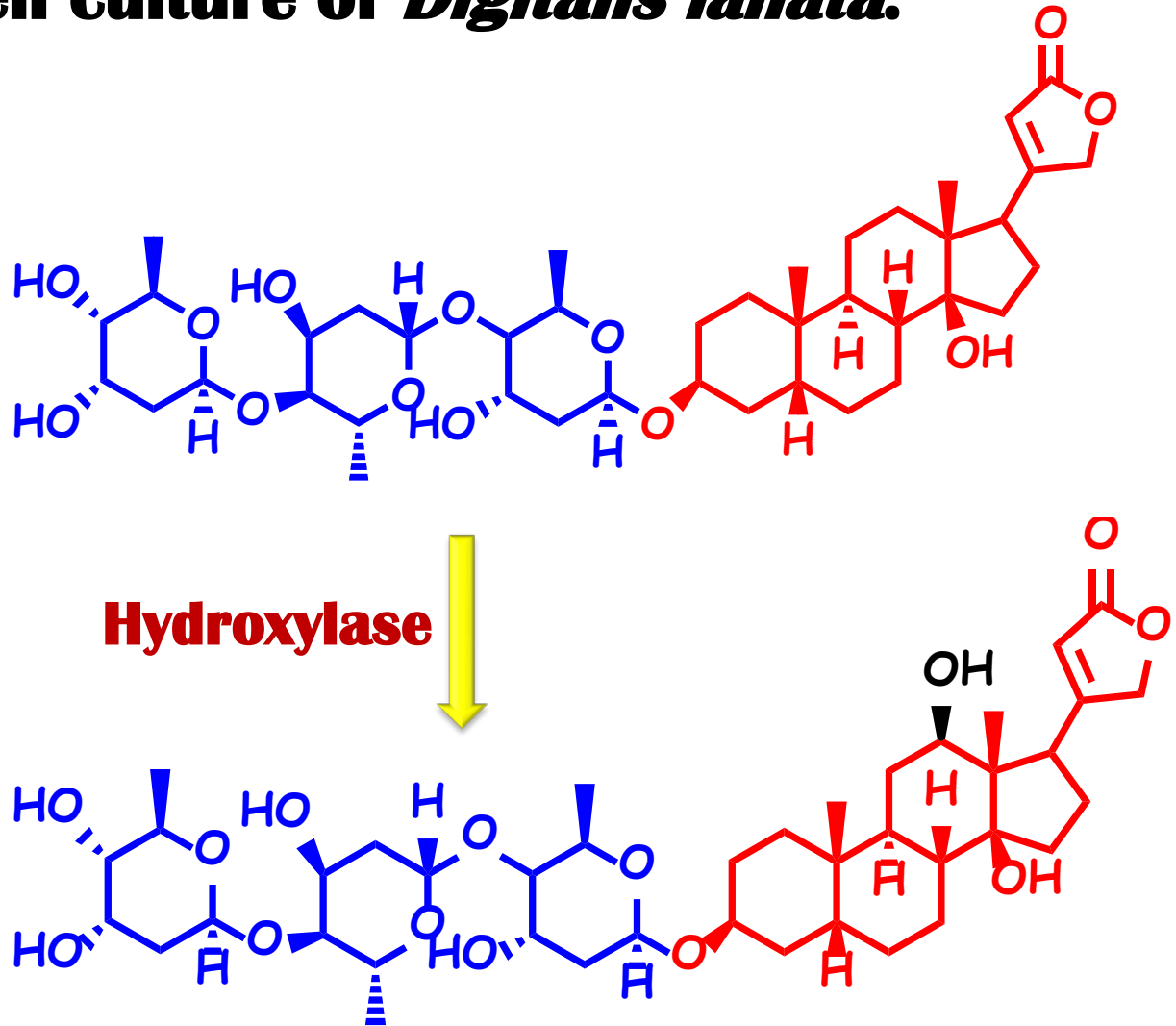
BIOTRANSFORMATION

- ❑ Using enzyme to transform less active compound to more active product. e.g. hydroxylation or glycosidation.



BIOTRANSFORMATION

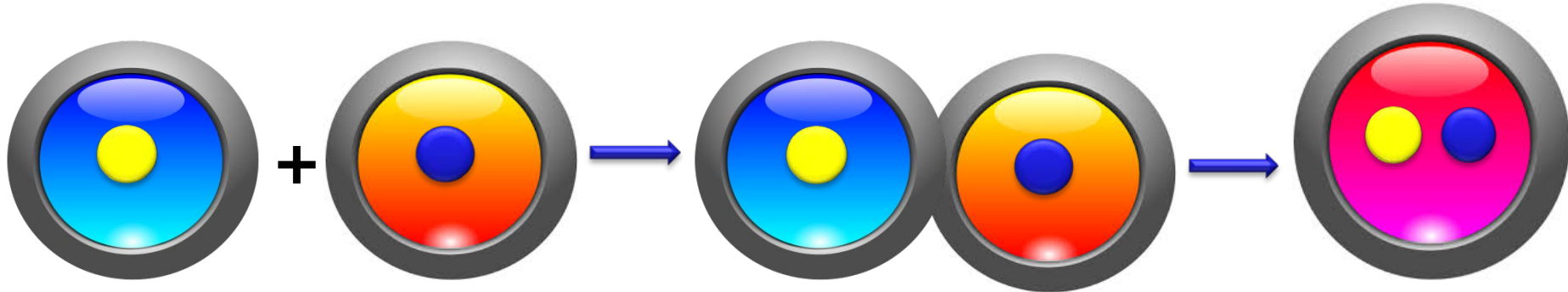
- ❑ Biotransformation of Digitoxin to digoxin by enzyme located in the cell culture of *Digitalis lanata*.



IMPROVEMENT OF PLANTS BY SOMATIC HYBRIDIZATION

Plant protoplasts

□ Ability of these Plant protoplasts to fuse with each other irrespective of their origin (Protoplast Fusion).



Spontaneous fusion.

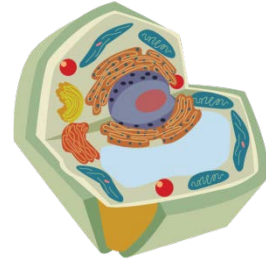
Induced fusion.

SOMATIC HYBRIDIZATION

1- ISOLATION OF PROTOPLAST.

a- Mechanically using a fine knife.

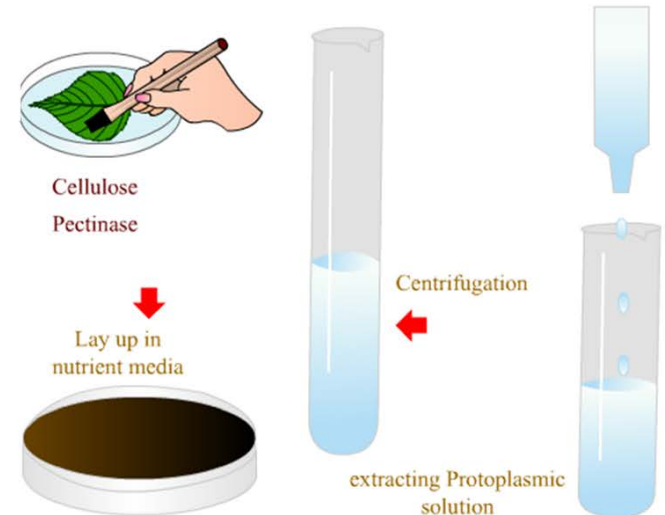
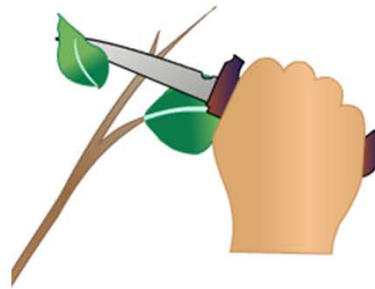
Cell Wall



b- The enzymatic isolation (Cellulase - pectinase).

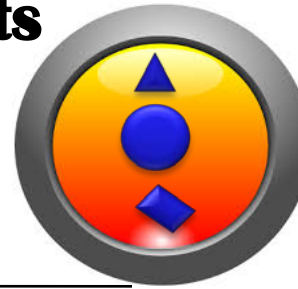
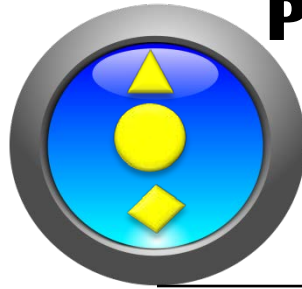


Sterile Leaf + Knife



HYBRIDS & CYBRIDS

Protoplasts



Nucleus



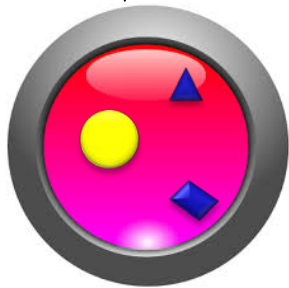
Mitochondria



Chloroplast



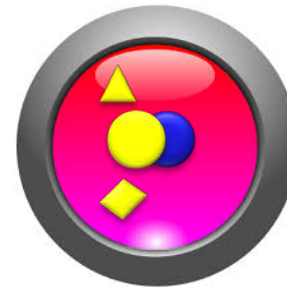
Heterokaryon



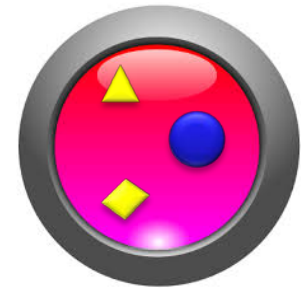
Cybrid



Hybrid



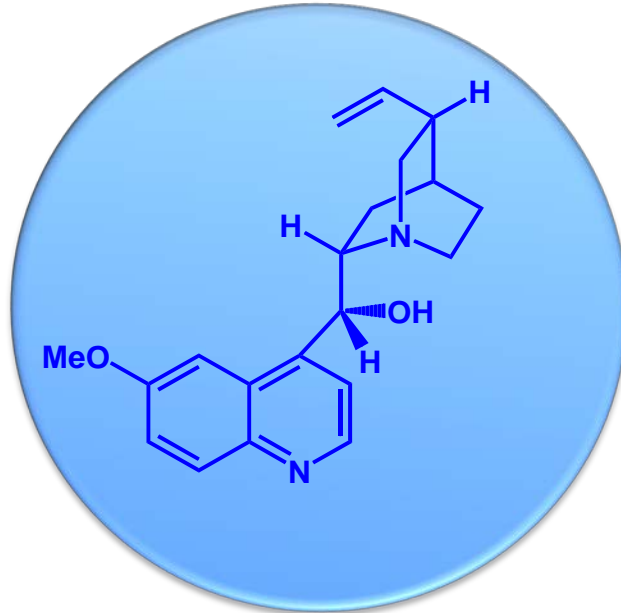
Hybrid



Cybrid

SOMATIC HYBRIDIZATION

- ❑ Valuable with species which are closely related.
- ❑ *Cinchona succirubra* (3% alkaloids) and *Cinchona ledgeriana* (5% alkaloids) form a hybrid (11 % alkaloids).



HAPLOID PLANTS



Culture in solid media



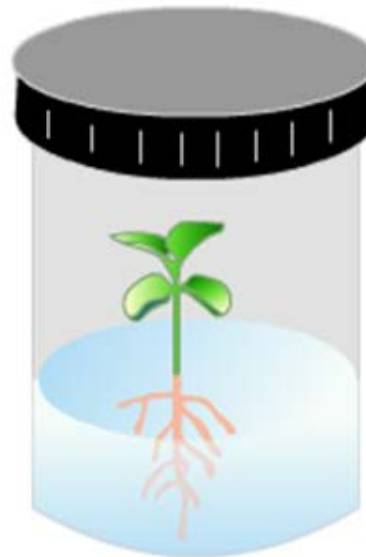
Selecting the
anthers



Remove the filament

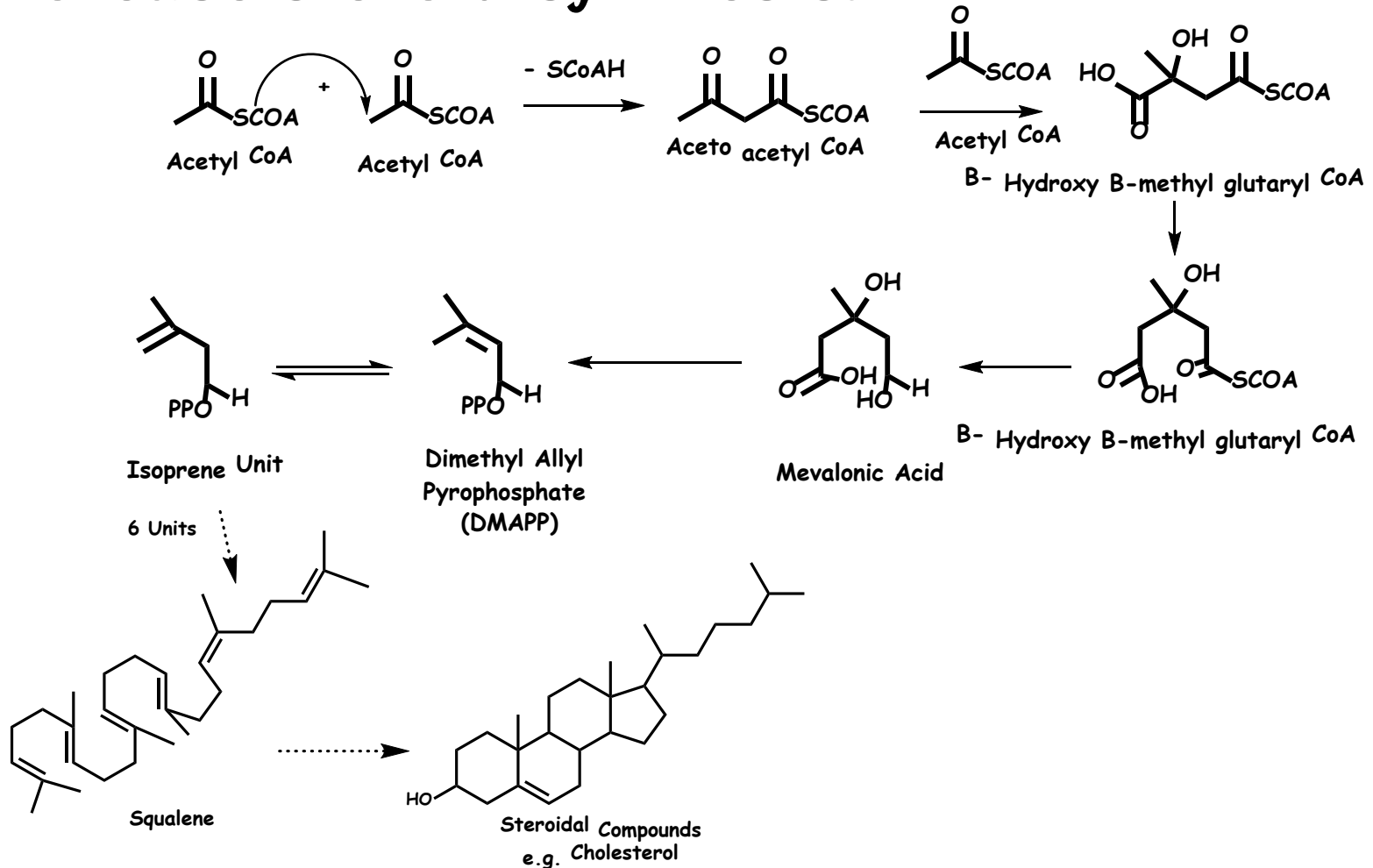


Culture in liquid
media



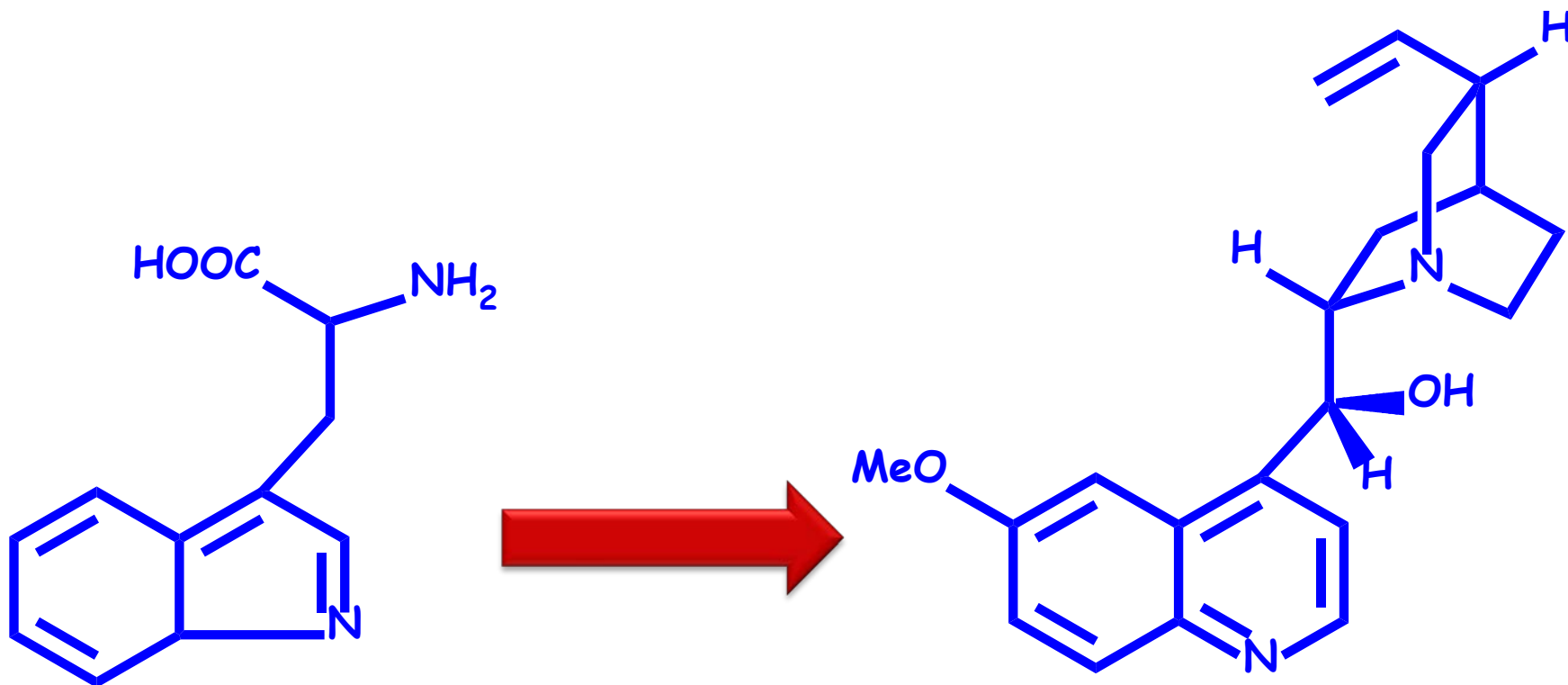
SECONDARY METABOLITES

- Addition of precursors e.g. Mevalonic acid increase steroid synthesis.



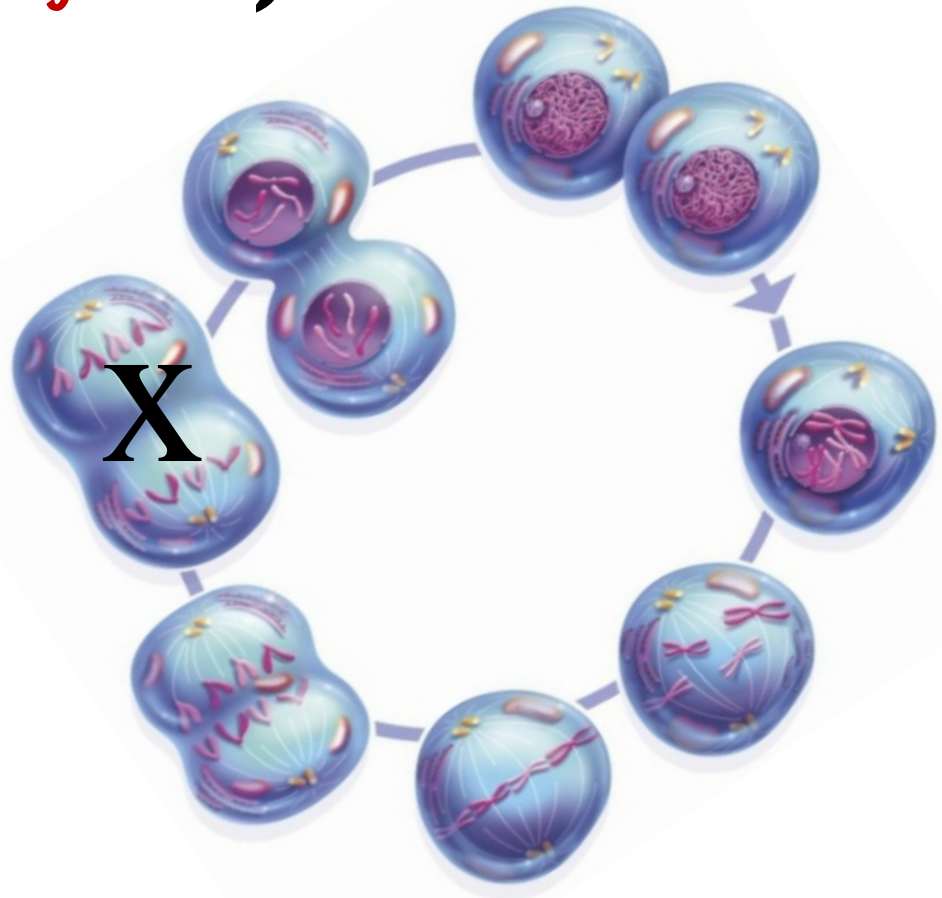
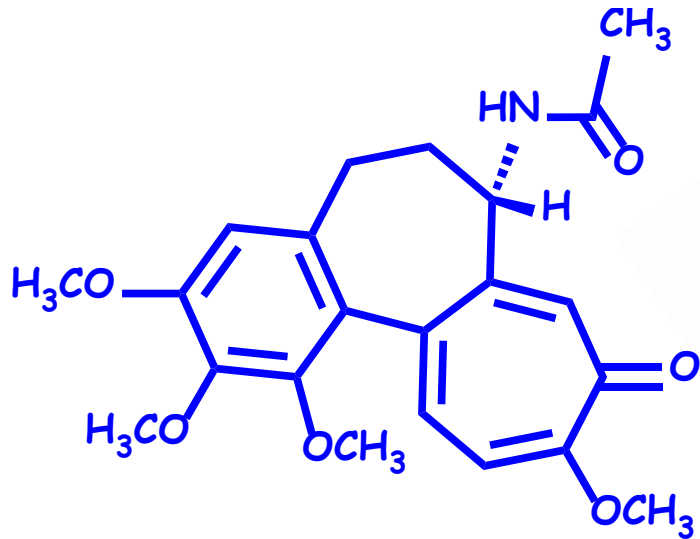
SECONDARY METABOLITES

- Addition of precursor (**L-tryptophan**) to the culture media of *Cinchona ledgeriana* increase its **quinoline alkaloid content**.



SECONDARY METABOLITES

- ❑ Addition of **colchicine** to cell suspension culture of *Valeriana spp.* increase the **valeprotriates** (**sixty fold**).





THANK



YOU

